

Variation in genome size and karyotype among closely related aphid parasitoids (Hymenoptera, Aphelinidae)

Vladimir E. Gokhman¹, Kristen L. Kuhn², James B. Woolley³, Keith R. Hopper²

1 Botanical Garden, Moscow State University, Moscow, Russia **2** Beneficial Insects Introduction Research Unit, ARS-USDA, 501 South Chapel Street, Newark, Delaware, United States of America **3** Department of Entomology, Texas A&M University, College Station, Texas, United States of America

Corresponding author: Keith R. Hopper (Keith.Hopper@ars.usda.gov)

Academic editor: M. Bressa | Received 25 October 2016 | Accepted 4 January 2017 | Published 23 February 2017

<http://zoobank.org/7D29D113-1F00-4445-80A2-6A2B3B4D30C1>

Citation: Gokhman VE, Kuhn KL, Woolley JB, Hopper KR (2017) Variation in genome size and karyotype among closely related aphid parasitoids (Hymenoptera, Aphelinidae). *Comparative Cytogenetics* 11(1): 97–117. <https://doi.org/10.3897/CompCytogen.v11i1.10872>

Abstract

Genome sizes were measured and determined for the karyotypes of nine species of aphid parasitoids in the genus *Aphelinus* Dalman, 1820. Large differences in genome size and karyotype were found between *Aphelinus* species, which is surprising given the similarity in their morphology and life history. Genome sizes estimated from flow cytometry were larger for species in the *A. mali* (Haldeman, 1851) complex than those for the species in the *A. daucicola* Kurdjumov, 1913 and *A. varipes* (Förster, 1841) complexes. Haploid karyotypes of the *A. daucicola* and *A. mali* complexes comprised five metacentric chromosomes of similar size, whereas those of the *A. varipes* complex had four chromosomes, including a larger and a smaller metacentric chromosome and two small acrocentric chromosomes or a large metacentric and three smaller acrocentric chromosomes. Total lengths of female haploid chromosome sets correlated with genome sizes estimated from flow cytometry. Phylogenetic analysis of karyotypic variation revealed a chromosomal fusion together with pericentric inversions in the common ancestor of the *A. varipes* complex and further pericentric inversions in the clade comprising *Aphelinus kurdjumovi* Mercet, 1930 and *Aphelinus hordei* Kurdjumov, 1913. Fluorescence *in situ* hybridization with a 28S ribosomal DNA probe revealed a single site on chromosomes of the haploid karyotype of *Aphelinus coreae* Hopper & Woolley, 2012. The differences in genome size and total chromosome length between species complexes matched the phylogenetic divergence between them.

Keywords

Aphelinidae, *Aphelinus*, parasitoid, genome size, flow cytometry, karyotype

Introduction

Genome size estimates and karyotypic studies provide data for comparative research at various taxonomic levels and allow evaluation of phylogenetic associations (Gokhman 2009, Hanrahan and Johnston 2011, Lopes et al. 2009). The completeness of genome assemblies can be difficult to assess, and independent estimates of genome size can aid in the assessment of completeness of genome assemblies (Gregory et al. 2013). Flow cytometry and Feulgen densitometry have been used to accurately measure genome size, and both methods have been extensively validated and various sources of error have been minimized through best-practice protocols (Gregory et al. 2013, Hare and Johnston 2011). Karyotypes can further help in assessing genetic linkage maps, and thus aid in mapping quantitative trait loci (Gokhman and Kuznetsova 2006). To visualize karyotypic features, various techniques of conventional and differential staining of chromosomes have been used, including fluorescence *in situ* hybridization (FISH), which allows physical mapping of DNA sequences onto chromosomes (Gadau et al. 2014, Macgregor and Varley 1988). Increasing the numbers of genome size estimates and karyotypes across the tree of life provides resources for the advancement of evolutionary genomics (Jacobson et al. 2013, Sharakhova et al. 2014). Furthermore, both flow cytometry and karyotypes can be used to detect cryptic species (Baur et al. 2014, Vergilino et al. 2012).

Genome size estimates have been published for more than 13,000 species of animals and plants (Animal Genome Size Database, <http://www.genomesize.com>; Plant DNA C-values Database, <http://data.kew.org/cvalues>; accessed 29 August 2014). There are currently 930 estimates of insect genome size in the Animal Genome Size Database, 152 of which are for species of Hymenoptera, and these genome sizes range from 98 to 1115 Mb. Genome size is usually considered constant within species, and limited intraspecific variation is a standard assumption in measurement and comparison of genome sizes. However, genome size can vary widely between closely related species (Gregory and Johnston 2008) and even within species (Biemont 2008, Bosco et al. 2007). The most common source of inter- and intraspecific genome size variation is differing amounts of repetitive DNA (Biemont 2008, Bosco et al. 2007). Differences in chromosome size can result from differences in heterochromatin content and amount of repetitive DNA in euchromatin, and differences in both chromosome size and number can result from fissions and fusions (Gokhman 2009, White 1973). Chromosome numbers and other karyotypic features have been published for about 70,000 species of plants and animals (Rice et al. 2014, White 1973; Tree of Sex: A database of sexual systems, doi: 10.1038/sdata.2014.15), including more than 1,500 species of Hymenoptera, whose haploid chromosome numbers range from 1 to 60 (Gokhman 2009, Ross et al. 2015).

Here we report genome size estimates and karyotypes for males and females in nine species of *Aphelinus* Dalman, 1820 (Hymenoptera: Chalcidoidea: Aphelinidae) all of which are parasitoids of aphids. Parasitoids are free-living as adults, but are

parasitic as larvae, and represent one of the most species-rich groups of insects, constituting more than 10% of all described insect species (Eggerton and Belshaw 1992, Heraty et al. 2007). Parasitoids are important regulators of arthropod populations, including major agricultural pests (Godfray 1994). The genus *Aphelinus* comprises more than 90 recognized species (Hopper et al. 2012; Universal Chalcidoidea Database, www.nhm.ac.uk/entomology/chalcidoids/index.html, accessed 10 October 2016). Within *Aphelinus*, several complexes of closely related species provide excellent opportunities to explore genetic differentiation, speciation, and the evolution of reproductive compatibility, host use, and morphology (Heraty et al. 2007, Hopper et al. 2012). We studied species in three complexes of *Aphelinus*: (1) five species in the *A. varipes* (Förster, 1841) complex from throughout Eurasia; (2) three species in the *A. mali* (Haldeman, 1851) complex from eastern Asia; (3) one species in the *A. daucicola* Kurdjumov, 1913 complex from North America. The *A. varipes* complex comprises 12 described species (Förster 1841, Hayat 1972, 1994, Hayat and Fatima 1992, Howard 1908, Kurdjumov 1913, Nikol'skaya and Yasnosh 1966, Pan 1992, Yasnosh 1963). The monophyly of the *A. varipes* complex is well supported by a combination of morphological and genetic characters (Heraty et al. 2007). However, some species within the complex show little morphological divergence, making identification difficult. The *A. mali* complex comprises 14 recognized species, some of which also show little morphological divergence (Ashmead 1888, Evans et al. 1995, Gahan 1924, Girault 1913, Haldeman 1851, Hayat 1998, Hopper et al. 2012, Prinsloo and Neser 1994, Timberlake 1924, Yasnosh 1963, Zehavi and Rosen 1988). The *A. daucicola* species complex comprises three species that differ from the members of the *A. mali* complex in several traits (Hopper et al. 2012). Using flow cytometry, we estimated the genome sizes of species in these complexes. We also made and examined chromosomal preparations to determine their karyotypes. We found consistent differences in genome size between complexes, and these differences correlated with differences in relative sizes estimated from karyotypes. We detected chromosomal rearrangements as well as karyotypic synapomorphies.

Materials and methods

Specimens

The parasitoid species studied, the sources of the colonies, and the permit and voucher numbers are listed in Table 1. These colonies were reared on aphids at the USDA-ARS, Beneficial Insect Introductions Research Unit, in Newark, Delaware, USA. Vouchers for these populations are maintained at -20 °C in 100% molecular grade ethanol at the Beneficial Insect Introduction Research Unit, Newark, Delaware. Females of the yellow-white strain of *Drosophila melanogaster* (Meigen, 1830) (stock number 1495, obtained from the Bloomington Drosophila Stock Center at

Table 1. The nine *Aphelinus* species studied, the year and country of their collection, permit and voucher numbers.

Species complex	Species	Authority	Year	Country	Permit and voucher
<i>A. varipes</i>	<i>A. atriplicis</i>	Kurdjumov, 1913	2000	Georgia	P526P-15-04274, VGg00_Dn
	<i>A. varipes</i>	(Förster, 1841)	2009	France	P526P-13-02503, VFr09_Rp
	<i>A. certus</i>	Yasnosh, 1963	2001	Japan	P526P-01-53096, VJp01_TU
	<i>A. kurdjumovi</i>	Mercet, 1930	2000	Georgia	P526P-13-02503, VGg00_Rp
	<i>A. hordei</i>	Kurdjumov, 1913	2011	France	P526P-15-04274, VFr11_Dn
<i>A. daucicola</i>	<i>A. daucicola</i>	Kurdjumov, 1913	2013	USA	P526P-15-04274, DUSA12_UD
<i>A. mali</i>	<i>A. glycinis</i>	Hopper et Woolley, 2012	2007	China	P526P-08-02142, MKor09_M
	<i>A. coreae</i>	Hopper et Woolley, 2012	2009	Korea	P526P-01-72318, MCh04_Bj
	<i>A. rhamni</i>	Hopper et Woolley, 2012	2005	China	P526P-01-53096, MCh05_Bj

Indiana University, <http://flystocks.bio.indiana.edu>) were used as internal controls for flow cytometry. All institutional and national guidelines for the care and use of laboratory animals were followed.

Flow cytometry

Live *Aphelinus* were sexed, flash frozen in liquid nitrogen, and stored at -80°C . To estimate genome sizes, we used the flow cytometry protocol described by Hanrahan and Johnston (2011) and Hare and Johnston (2011). We dissected heads from both males and females of the *Aphelinus* species in cold Galbraith buffer (Galbraith et al. 1983). Heads of female *D. melanogaster* were used as internal standards (1C = 175 Mb or 0.17 pg). To release the nuclei from cells, heads from 15 female *Aphelinus* and one female *Drosophila* Fallén, 1823 for each replicate were ground together in one milliliter of cold Galbraith buffer using 15 strokes of the “A” pestle in a 2-ml Kontes Dounce tissue grinder. As with other Hymenoptera, *Aphelinus* species have haplodiploid sex determination, with males coming from unfertilized eggs and females from fertilized eggs. Thus males carried half as much DNA per cell as females, which made male genome sizes too close to that of *D. melanogaster*. Thus when processed the heads of 15 males per replicate as described above, we included the heads from 15 females of the same parasitoid species as internal standards. The samples were passed through a 35 micron filter and then stained with 40 parts per million of propidium iodide in the dark for 3–5 hours at 4°C . Samples were analyzed with laser excitation at 488 nm on a Becton Dickinson FACSCalibur Flow Cytometer (BD Biosciences, San Jose, CA, USA) at CTCR Core Facility, University of Delaware. Red fluorescence from the propidium iodide was detected using an FL2 filter. Three to six replicates were measured for females and males of each species.

The haploid content of DNA in megabases (Mb) was calculated for each *Aphelinus* sample from the ratio of mean fluorescence of the sample to mean fluorescence of the

standard times the genome size of the standard. We report genome size estimates in megabases, but also give estimates in picograms (pg) calculated by dividing the amount of DNA in Mb by the standard 1C value of 978 Mb.

Karyotypes

Chromosome preparations were made from cerebral ganglia of prepupae using a modified version of the technique in Imai et al. (1988). Wasps were dissected in 0.5% hypotonic sodium citrate solution containing 0.005% colchicine, and the tissues were incubated in fresh solution for ~30 minutes at room temperature. The material was transferred to a pre-cleaned microscope slide using a Pasteur pipette and gently flushed with Fixative I (glacial acetic acid: absolute ethanol: distilled water 3:3:4). Tissues were disrupted in an additional drop of Fixative I using dissecting needles. Another drop of Fixative II (glacial acetic acid: absolute ethanol 1:1) was then applied to the center of the area and blotted off the edges of the slide. The slide was air dried for ~30 minutes at room temperature. For conventional staining, preparations were stained with freshly prepared 3% Giemsa solution in 0.05M Sørensen's phosphate buffer ($\text{Na}_2\text{HPO}_4 + \text{KH}_2\text{PO}_4$, pH 6.8). Mitotic divisions were studied and photographed using an optic microscope Zeiss Axioskop 40 FL fitted with a digital camera AxioCam MRc (Carl Zeiss, Oberkochen, Germany). To obtain karyograms, the resulting images were processed with image analysis programs: Zeiss AxioVision version 3.1 and Adobe Photoshop version 8.0. Mitotic chromosomes were measured for 5–19 cells in 1–6 wasps per species using Adobe Photoshop. We report total length (μm) of all chromosomes in each karyotype for males and females; for diploid sets, we divided total length by two to make the values comparable to haploid sets. We also report relative lengths (RL: $100 \times \text{length of each chromosome divided by total length of the set}$) and centromeric indices (CI: $100 \times \text{length of shorter arm divided by total length of a chromosome}$) for females of each species. Chromosomes were classified into metacentric (M) or acrocentric (A) according to the guidelines in Levan et al. (1964).

Fluorescence *in situ* hybridization

A custom biotinylated fragment from the 28S rDNA gene was used to probe *A. coreae* chromosomes with fluorescence *in situ* hybridization (FISH). To prepare the probe, we extracted DNA from ~50 adult parasitoids using a Qiagen DNeasy Blood and Tissue Kit (Qiagen, Valencia, CA, USA). From this DNA, we amplified a ~650 nt fragment of the 28S rDNA gene using the following primers and PCR protocol: reaction mix - 5 μl NEB PCR buffer and 0.5 μl *Taq* polymerase (New England Biolabs, Ipswich, MA, USA), 4 μl each of 2.5 mM dATP, dCTP, dGTP, 4 μl 0.25 mM dTTP plus 1 μl 1mM biotinylated-11-dUTP, 1 μl 10 μM forward primer (5'-cgt gtt gct tga tag tgc agc) and 1 μl 10 μM reverse primer (5'-tca aga cgg gtc ctg aaa gt), 4 μl genomic DNA (50 ng/ μl),

21.5 µl ultrapure H₂O; cycling - 3 min at 95 °C, then 35 cycles 95 °C for 30 sec, 55°C for 30 sec, and 72°C for 60 sec, and a final extension at 72 °C for 3 min. Unincorporated dNTPs, primers, and other unwanted components were removed from the PCR product using precipitation with sodium acetate and ethanol, and the resulting pellet was resuspended in 50 µl ultrapure H₂O, yielding a solution of probe at 300 ng/µl.

Chromosomes were prepared for probing using the protocol described above for karyotyping. To probe the chromosomes, a protocol modified from Matsumoto et al. (2002) was used. Chromosomes were baked onto slides at 65°C and UV crosslinked in a Spectrolinker XL-1000 UV Crosslinker (Spectronics, Westbury, NY, USA) twice at 120 mJ/cm². The slides were treated with RNase A, dehydrated with ethanol, denatured in formamide, and then dehydrated again with ethanol. The slides were then treated with proteinase K and dehydrated a third time with ethanol. Hybridization solution was prepared from the biotinylated probe (7 µg in 23 µl), formamide (90 µl), 30% dextran sulfate solution (60 µl), 10 mg/ml salmon sperm DNA (5 µl), and 20x SSC (22 µl). This solution was denatured at 95°C, placed on ice, and then 50 µl was applied to each slide, which were then covered with parafilm and left in a moist chamber at 37°C for 12–16 h. The slides were washed twice in formamide (50% in 2x SSC) and twice in 2x SSC with gentle shaking, and transferred to BN buffer (100 mM NaHCO₃, 0.1% Nonidet P-40) for 10 min at room temperature. After this, blocking buffer (100 mM NaHCO₃, 0.05% Nonidet P-40, 0.02% NaN₃, 5% non-fat dry milk) was applied, and the slides were covered with parafilm and incubated for 10 min at room temperature. The buffer was removed, streptavidin-Alexa fluor 568 conjugate (Thermo Fisher Scientific, Waltham, MA, USA), diluted 1/50 in blocking buffer, was added, and the slides were covered with parafilm and incubated at 37°C for 1 h. The slides were washed with three changes of BN buffer in a light-tight chamber with gentle shaking. Signal enhancement was done following Pinkel et al. (1986). Fifty microliters of biotinylated goat anti-Avidin D (Vector Laboratories, Burlingame, CA, USA), diluted 1/50 in the blocking buffer, was applied to each slide, which were then covered with parafilm and incubated at 37°C for 1 hour. The slides were then washed with BN buffer, more streptavidin-labeled fluor was added, the slides were washed again with BN buffer, and then air-dried in the dark at room temperature. Anti-fade medium (ProLong Gold Antifade Reagent with DAPI, Cell Signaling Technology, Danvers, MA, USA) was added and a glass coverslip was placed over the chromosomal preparation. The chromosomes were imaged using a Zeiss 510 NLO Multiphoton microscope and a Zeiss Elyra PS 1 microscope (Carl Zeiss, Pleasanton, CA, USA) with confocal microscopy at the Bio-Imaging Center, Delaware Biotechnology Institute, Newark, DE, USA.

Data analysis

Genome sizes and total lengths of chromosome sets were compared among species and between sexes in generalized linear models with species and sex as fixed main-effects

and Poisson error distributions using the `glm` function in R (R Core Team 2014). The set of relative lengths among species in a multivariate analysis of variance was compared with the Pillai–Bartlett statistic and the `manova` function in R. Centromeric indexes among species in generalized linear models were compared for each chromosome with species as a fixed effect and Poisson error distributions using the `glm` function in R. Because chromosomal formulae were different for the *A. varipes* complex versus the *A. mali* and *A. daucicola* complexes, we analyzed the effects of species on relative lengths and centromeric indexes separately within these groups. For genome size, the experimental unit was either 15 heads of female parasitoids and one *D. melanogaster* head pooled or 15 heads of male parasitoids and 15 heads of female parasitoids pooled. For total lengths of chromosome sets and relative lengths and centromeric indexes of chromosomes, the experimental unit was an individual mitotic cell. Post-hoc comparisons of means were done using the `glht` and `cld` functions in the `multcomp` package in R. We tested the relationships between genome sizes from flow cytometry and total lengths of chromosome sets with linear regression using the `lm` function in R. Data are archived on the Ag Data Commons website (data.nal.usda.gov; DOI 10.15482/USDA.ADC/1329930).

Results

Genome sizes from flow cytometry

Haploid genome sizes of *Aphelinus* differed among species (model deviance = 444.0; residual deviance = 5.4; $df = 6, 60$; $P < 0.0001$). Female genome sizes ranged from 330 to 483 Mb so the largest was 1.5 times the smallest (Table 2, which also shows results of multiple comparisons among means of each species). Female and male *Aphelinus* had similar haploid genome sizes, with female and male sizes within 2–13 Mb (1–4 percent) of one another, so the sexes did not differ significantly (model deviance = 1.1; residual deviance = 4.3; $df = 1, 59$; $P = 0.30$). Genomes (averaged across sexes) in the *A. mali* complex were significantly larger (37–148 Mb or 9–44 percent) than those *A. varipes* complex, and genomes in the *A. varipes* complex were significantly larger (1–59 Mb or 1–18 percent) than those in the *A. daucicola* complex (model deviance = 378.10; residual deviance = 73.3; $df = 6, 60$; $P < 0.0001$). The genome of *A. rhamni* was significantly larger (43–53 Mb or 10–12 percent) than the genomes of the other species in the *A. mali* complex. The genome of *A. hordei* was significantly larger (40–58 Mb or 9–17 percent) than the genomes of the other species in the *A. varipes* complex.

Karyotypes

Species in the *A. varipes* complex had four chromosomes in haploid males and thus eight chromosomes in diploid females, whereas species in the *A. mali* and *A. daucicola*

Table 2. Haploid genome sizes of nine *Aphelinus* species estimated from flow cytometry. Shared letters after means indicates that they do not differ significantly.

Species complex	Species	Sex	n replicates	Genome size		95% CI
				(pg)	(Mb)	(Mb)
<i>A. varipes</i>	<i>A. atriplicis</i>	female	6	0.361	353a	338–368
		male	3	0.366	358a	337–380
	<i>A. varipes</i>	female	3	0.340	333a	313–354
		male	3	0.348	340a	320–362
	<i>A. certus</i>	female	6	0.369	361a	347–377
		male	3	0.375	367a	346–390
	<i>A. kurdjumovi</i>	female	4	0.356	348a	331–367
		male	3	0.363	355a	334–377
	<i>A. hordei</i>	female	4	0.402	393b	374–412
		male	3	0.406	397b	375–421
<i>A. daucicola</i>	<i>A. daucicola</i>	female	4	0.337	330a	313–348
		male	3	0.351	343a	322–364
<i>A. mali</i>	<i>A. glycinis</i>	female	6	0.442	432c	416–449
		male	5	0.441	431c	413–450
	<i>A. coreae</i>	female	3	0.449	439c	416–464
		male	3	0.454	444c	421–468
	<i>A. rhamni</i>	female	6	0.494	483d	466–501
		male	3	0.498	487d	463–513

complexes had five chromosomes in haploids and thus ten chromosomes in diploids (Table 3; Figs 1–2). Karyotypes of the *A. varipes* complex usually included a large metacentric chromosome 1 and a small metacentric chromosome 2 and small acrocentric chromosomes 3 and 4, except for *A. kurdjumovi*, in which the small metacentric chromosome 2 appears to have been replaced by an acrocentric chromosome 2 of similar size; whereas species in the *A. mali* and *A. daucicola* complexes had metacentric chromosomes only, and their chromosomes showed a continuous gradation in length (Tables 4 and 5). Relative lengths of chromosome sets differed significantly among species in the *A. varipes* complex ($F = 3.2$; $df = 16, 540$; $P < 0.0001$), but did not quite differ significantly among species in the *A. mali* and *A. daucicola* complexes ($F = 1.7$; $df = 12, 255$; $P = 0.07$).

Centromeric indexes for chromosome 1 did not differ among species, and centromeric indexes for chromosome 2 did not differ among species in the *A. mali* and *A. daucicola* complexes. However, in the *A. varipes* complex, the centromeric index of chromosome 2 in *A. hordei* was significantly lower than in other members of the *A. varipes* complex. Centromeric indexes for chromosomes 3 and 4 in *A. daucicola* were significantly lower than those for *A. rhamni*, and the centromeric index for chromosome 5 in *A. daucicola* was significantly lower than those for *A. coreae* and *A. rhamni* (Table 6). Total lengths of chromosome sets differed among species

Table 3. Karyotypic features of nine *Aphelinus* species. Shared letters after means indicate that they do not differ significantly within each sex.

	species complex	species	number nuclei measured	number chromosomes	chromosomal formula	total length of chromosome set (μm)	
						mean	95% confidence interval
female	<i>A. varipes</i>	<i>A. atriplicis</i>	14	8	4M + 4A	15.6ac	13.7–17.9
		<i>A. varipes</i>	16	8	4M + 4A	15.1ab	13.3–17.1
		<i>A. certus</i>	19	8	4M + 4A	14.0a	12.4–15.8
		<i>A. kurdjumovi</i>	13	8	2M + 6A	16.8ac	14.8–19.2
		<i>A. hordei</i>	8	8	4M + 4A	14.3ab	11.9–17.1
	<i>A. daucicola</i>	<i>A. daucicola</i>	15	10	10M	16.8ac	14.8–19.0
	<i>A. mali</i>	<i>A. glycinis</i>	5	10	10M	19.6ac	16.1–23.9
		<i>A. coreae</i>	6	10	10M	21.3c	17.9–25.4
		<i>A. rhamni</i>	19	10	10M	18.4bc	16.5–20.4
male	<i>A. varipes</i>	<i>A. atriplicis</i>	3	4	2M + 2A	16.3ab	12.3–21.6
		<i>A. certus</i>	3	4	2M + 2A	25.0bc	19.9–31.3
		<i>A. hordei</i>	21	4	2M + 2A	17.8b	16.1–19.7
	<i>A. daucicola</i>	<i>A. daucicola</i>	7	5	5M	23.7ac	20.4–27.6
	<i>A. mali</i>	<i>A. coreae</i>	6	5	5M	29.8c	25.8–34.5
		<i>A. rhamni</i>	4	5	5M	22.5bc	18.3–27.7

M = metacentric; A = acrocentric.

Table 4. Relative lengths of chromosomes in *Aphelinus* species. Means with 95% confidence intervals in parentheses.

Species complex	Species	Chromosome				
		1	2	3	4	5
		Relative length				
<i>A. varipes</i>	<i>A. atriplicis</i>	40	26	18	16	
		(38–42)	(24–28)	(17–20)	(14–17)	
	<i>A. varipes</i>	41	26	18	15	
		(39–43)	(24–27)	(17–19)	(14–17)	
	<i>A. certus</i>	40	26	18	16	
		(38–43)	(24–28)	(17–20)	(14–17)	
	<i>A. kurdjumovi</i>	43	24	18	15	
		(40–45)	(22–26)	(17–20)	(14–17)	
	<i>A. hordei</i>	41	25	19	16	
		(37–44)	(23–27)	(17–21)	(14–18)	
<i>A. daucicola</i>	<i>A. daucicola</i>	24	22	19	18	18
		(23–26)	(20–24)	(18–21)	(17–20)	(17–20)
<i>A. mali</i>	<i>A. glycinis</i>	24	22	21	18	18
		(21–27)	(19–25)	(18–24)	(16–21)	(16–21)
	<i>A. coreae</i>	24	22	20	18	18
		(21–27)	(20–25)	(18–23)	(16–21)	(16–21)
	<i>A. rhamni</i>	24	22	20	18	18
		(23–26)	(21–24)	(19–22)	(17–19)	(17–19)



Figure 1. Haploid mitotic karyograms of six *Aphelinus* species. **a** *A. atroplicis* **b** *A. certus* **c** *A. hordei* **d** *A. coreae* **e** *A. rhamni* **f** *A. daucicola*. Species in the *A. varipes* complex have $n = 4$ versus $n = 5$ in the *A. mali* and *A. daucicola* complexes. Scale bar: 10 μm .

(model deviance = 65.1; residual deviance = 153.2; $df = 8, 150$; $P < 0.0001$) and between sexes (model deviance = 34.3; residual deviance = 118.9; $df = 1, 149$; $P < 0.0001$). Total lengths ranged from 14 to 21 μm so the longest set was 1.5 times the shortest (Table 3). Total lengths were significantly greater in the *A. mali* complex than in the *A. varipes* complex for both males and females, with the values in *A. daucicola* complex intermediate between these extremes (females: model deviance = 20.5; residual deviance = 92.8; $df = 2, 112$; $P < 0.0001$; males: model deviance = 26.2; residual deviance = 38.8; $df = 2, 41$; $P < 0.0001$). Mean total chromosome length correlated with mean genome size estimated from flow cytometry ($F = 6.3$;

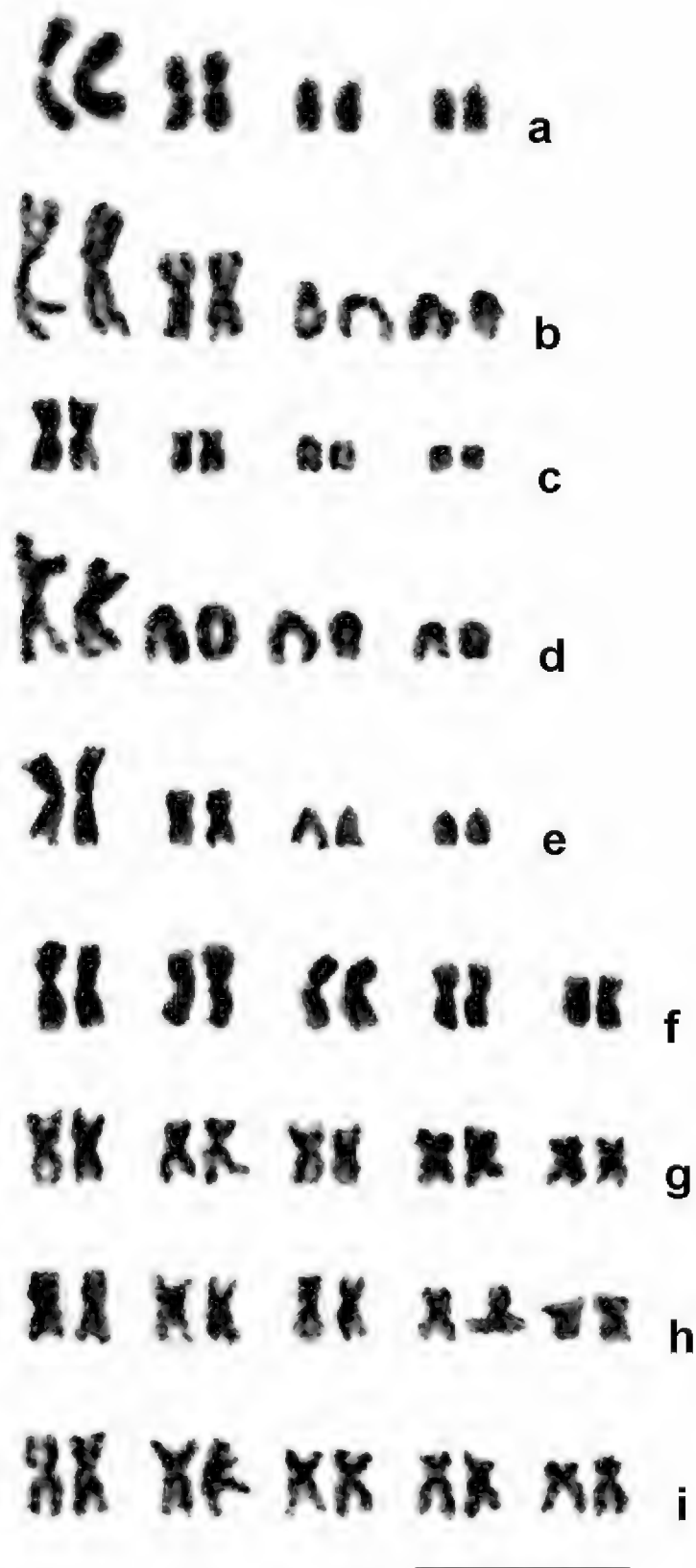


Figure 2. Diploid mitotic karyograms of nine *Aphelinus* species. **a** *A. atriplicis* **b** *A. certus* **c** *A. hordei* **d** *A. kurdjumovi* **e** *A. varipes* **f** *A. coreae* **g** *A. glycinis* **h** *A. rhamni*, **i** *A. daucicola*. Species in the *A. varipes* complex have $2n = 8$ versus $2n = 10$ in the *A. mali* and *A. daucicola* complexes. Scale bar: 10 μm .

$df = 1, 7$; $P = 0.04$; $r^2 = 0.47$; Fig. 3), primarily because of the difference in total chromosome length between the *A. mali* and *A. varipes* complexes.

Hybridization with a 28S rDNA probe revealed a single rDNA cluster on chromosomes of the haploid set and two rDNA clusters in the diploid set (Fig. 4). These clusters were near the centromere on a medium-sized metacentric chromosome.

Table 5. Centromeric indexes of chromosomes in *Aphelinus* species. Means (95% confidence intervals); shared letters within a species complex and chromosome indicate means that are not significantly different.

complex	species	Chromosome				
		1	2	3	4	5
		Centromeric index				
<i>A. varipes</i>	<i>A. atriplicis</i>	46a	47b	0	0	
		(44–49)	(45–50)			
	<i>A. varipes</i>	46a	47b	0	0	
		(44–49)	(45–49)			
	<i>A. certus</i>	46a	46ab	0	0	
		(44–49)	(44–48)			
	<i>A. kurdjumovi</i>	47a	0	0	0	
		(44–49)				
<i>A. mali</i> and <i>A. daucicola</i>	<i>A. daucicola</i>	45a	45a	41a	39a	38a
		(43–48)	(43–48)	(38–43)	(37–41)	(36–40)
	<i>A. glycinis</i>	48a	44a	44ab	44ab	42ab
		(43–52)	(40–48)	(40–48)	(40–49)	(39–47)
	<i>A. coreae</i>	44a	47a	43ab	44ab	44b
		(41–48)	(43–51)	(40–47)	(40–47)	(40–47)
	<i>A. rhamni</i>	46a	46a	45b	45b	43b
		(44–49)	(44–48)	(43–47)	(43–47)	(41–45)

Table 6. Analysis of deviance for differences in centromeric indexes among species of *Aphelinus*; acrocentric chromosomes were not included in these analyses.

complex	chromosome	model		residual		
		df	deviance	df	deviance	P
<i>A. varipes</i>	1	4	0.2	135	23.1	1.00
	2	3	9.3	110	22.3	0.03
<i>A. mali</i> and <i>A. daucicola</i>	1	3	1.8	86	20.7	0.62
	2	3	1.2	86	23.0	0.74
	3	3	7.2	86	33.2	0.07
	4	3	16.6	86	29.0	0.0008
	5	3	12.7	86	52.8	0.005

Discussion

The large genome size differences between the *A. varipes* complex versus the *A. mali* complex matched the phylogenetic divergence between these complexes (Heraty et al. 2007). The difference between the complexes is also supported by the difference in karyotypes: four chromosomes with one-two metacentrics and two-three acrocentrics in the *A. varipes* complex versus five metacentric chromosomes in the *A. mali* complex.

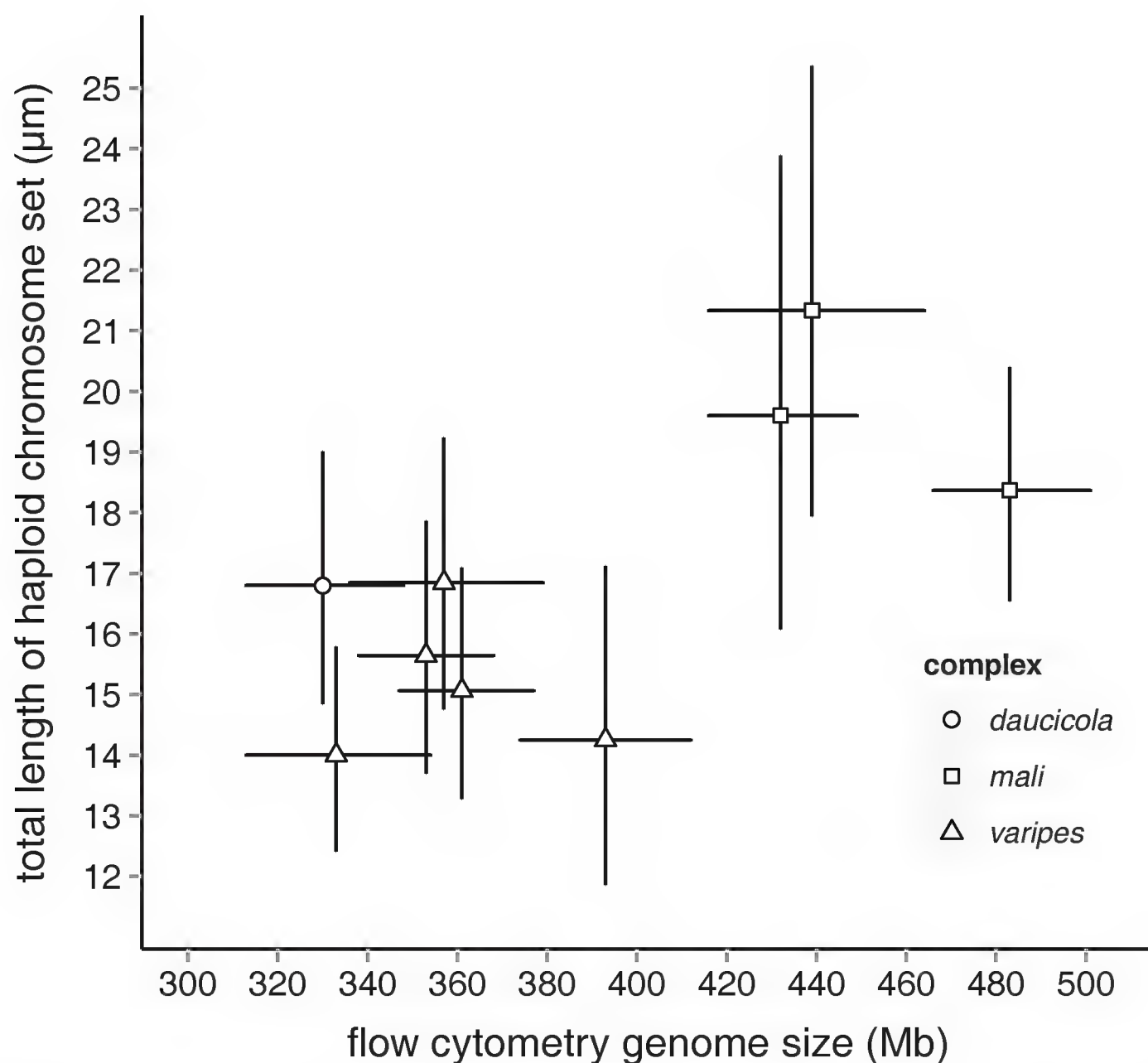


Figure 3. Total length of chromosome set (μm) versus flow cytometry genome size (Mb) for nine *Aphelinus* species. Error bars are 95% confidence intervals.

However, the additional chromosome in the *A. mali* complex does not necessarily account for a corresponding increase in genome size, because *A. daucicola* also has five chromosomes and yet has the smallest genome we observed. Four haploid genome sizes have been published for the family Aphelinidae, including an estimate of 635 Mb (=0.65 pg) for *Aphelinus abdominalis* (Dalman, 1820) (Ardila-Garcia et al. 2010) which is much larger than our estimates for other species of *Aphelinus*. However, beyond *Aphelinus*, other Hymenoptera genera have genome sizes spanning wide ranges (e.g., some ants, *Tapinoma* Förster, 1850, 362–597 Mb; *Solenopsis* Westwood, 1840, 372–753 Mb; <http://www.genomesize.com>). This also applies to some genera of parasitoids, e.g. *Leptopilina* Förster, 1869 (Figitidae) with genome sizes ranging from 363 to 520 Mb (Gokhman et al. 2011).

Genome size has been hypothesized to depend on several factors, including eusociality, parasitism, and developmental biology in insects (Ardila-Garcia and Gregory 2009, Gregory 2002, Johnston et al. 2004). These hypotheses come from the asso-

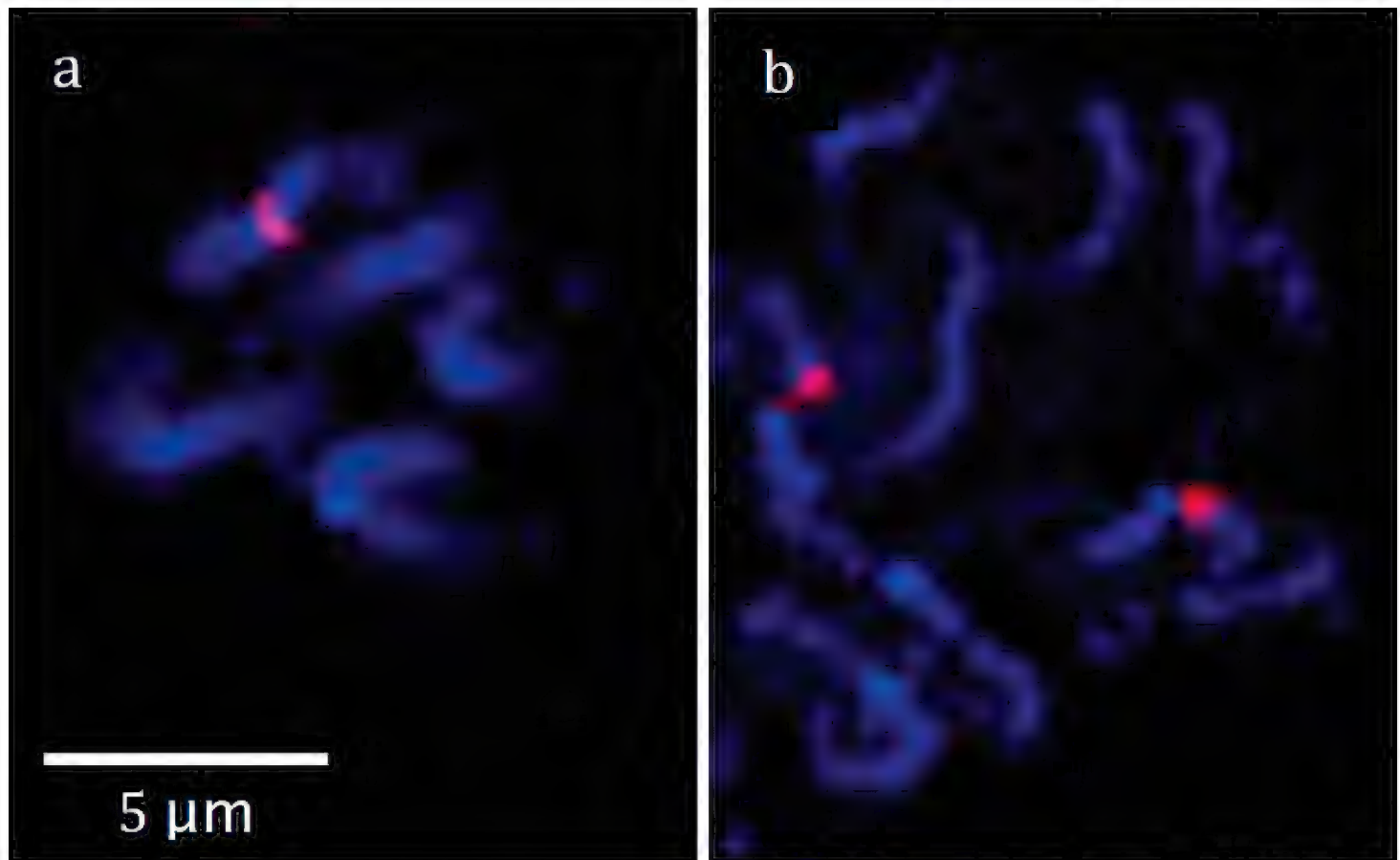


Figure 4. Fluorescence *in situ* hybridization with 28S rDNA probe. **a** metaphase chromosomes of the haploid karyotype and **b** prometaphase chromosomes of the diploid karyotype of *A. coreae*. Red = hybridization signal (a single rDNA cluster in the haploid set and paired clusters in the diploid set), blue = counterstaining of chromosomes with DAPI.

ciations between genome size, cell size, and cell division rates found in many taxa (Ardila-Garcia and Gregory 2009, Gregory 2005). The mean genome size for species of parasitic Hymenoptera (293 Mb) does not differ greatly from the mean genome size for species of eusocial Hymenoptera (333 Mb), but the genomes for both groups are significantly smaller than those for species of non-parasitoid solitary Hymenoptera (469 Mb) (Ardila-Garcia and Gregory 2009). However, it is unclear why there should be so much variation in genome size among species of *Leptopilina* or *Aphelinus*, given the very similar biologies within each genus.

Mapping karyotypic data on a molecular phylogeny of *Aphelinus* and two outgroup species allowed reconstruction of karyotype evolution in the species we studied (Fig. 5). The phylogeny was modified from Heraty et al. (2007) with results from Heraty et al. (2013), Kim and Heraty (2012), and unpublished data. Chromosomal formulae were mapped on the phylogeny using Mesquite (Maddison and Maddison 2016). Chromosomal formulae for *Aphytis mytilaspidis* (Le Baron, 1870) and *Encarsia formosa* Gahan, 1924 are from (Gokhman 2003). Concerning chromosome number, *Aphelinus asychis* Walker, 1839, has four chromosomes (Gokhman 2003), which is what we found for all the species we studied in the *A. varipes* complex. On the other hand, we found five chromosomes for *A. daucicola* and three species in the *A. mali* complex, which is also the chromosome number reported for *Aphelinus mali* (Haldeman, 1851) (Viggiani 1967).

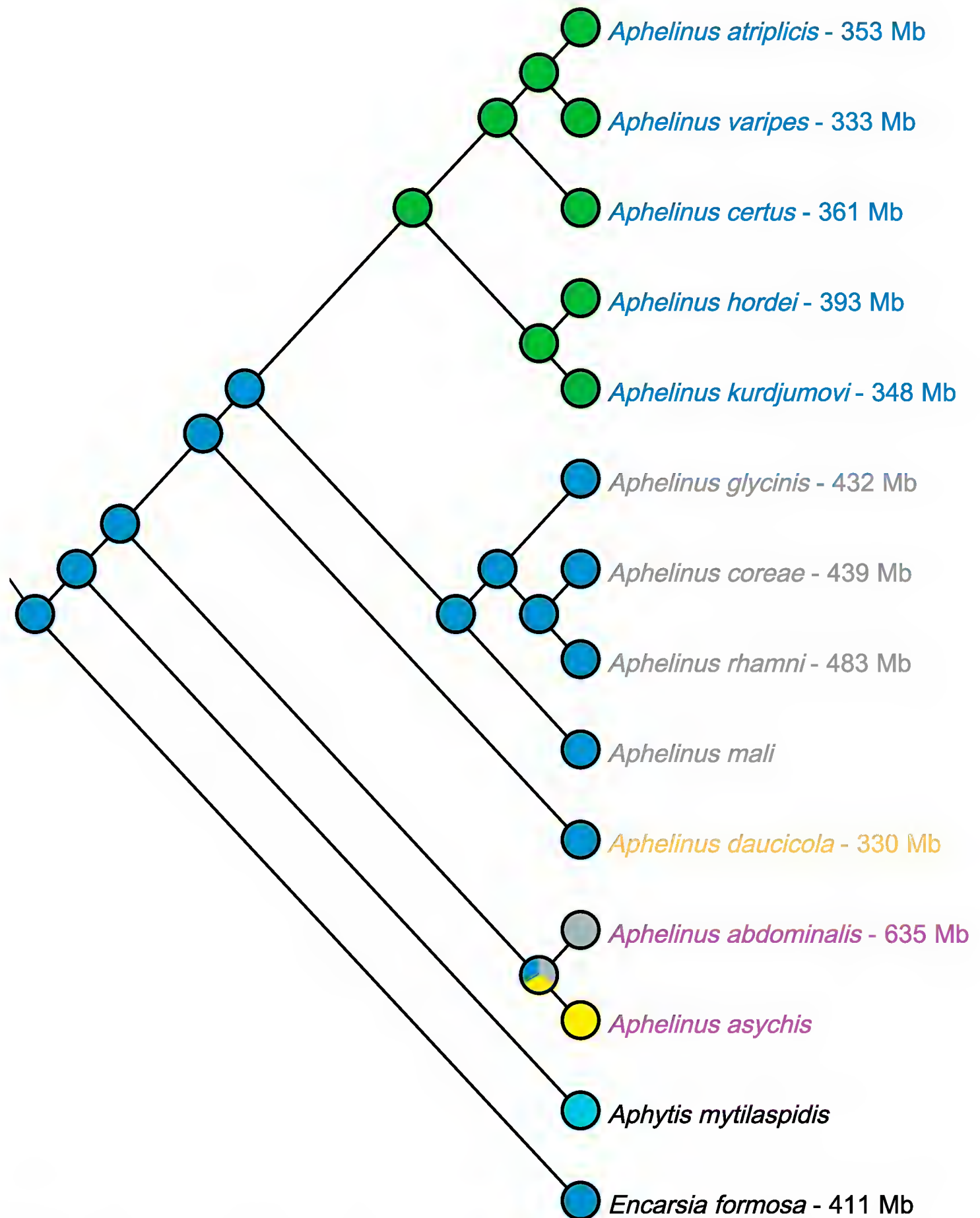


Figure 5. Chromosomal formulae and genome sizes on phylogeny of *Aphelinus* species and several outgroups. Blue = 5 metacentric chromosomes; aqua = 1 metacentric and 4 acrocentric chromosomes; green = 2 metacentric and 2 acrocentric chromosomes; dark green = 1 metacentric and 3 acrocentric chromosomes; yellow = 2 metacentric, 1 subtelocentric, and 1 acrocentric chromosome; grey = unknown. Numbers after species names are genome sizes estimated from flow cytometry; values for *Aphelinus abdominalis* and *E. formosa* are from (Ardila-Garcia et al. 2010). The different colors for species of *Aphelinus* indicate membership in the four species complexes for which data are available.

The *A. varipes* and *A. mali* complexes are sister clades, but *A. asychis* is the most basal species in the phylogeny of the genus, at least for the species for which there are phylogenetic data. This information alone would suggest four chromosomes is the ancestral state. However, five or six chromosomes are the numbers most frequently reported in chalcidoids (although it has been hypothesized that 9–10 is the ancestral state) (Gokhman 2009). Furthermore, five chromosomes has been reported for *Aphytis mytilaspidis* (Rössler and DeBach 1973), a species in the genus most closely related to *Aphelinus* for which chromosome number has been reported, and five chromosomes have been reported for many species of *Encarsia* Förster, 1878, a genus of aphelinids in another subfamily of Aphelinidae (Gokhman 2009). Chromosomal fusion (and hence decreased chromosome number) is a trend of karyotype evolution in many groups of organisms, including parasitic Hymenoptera (Gokhman 2009). Chromosomal fissions are also possible, but they are substantially less frequent, probably because they break existing linkage groups and therefore can decrease fitness. Thus, the reduced chromosome number in *A. asychis* has probably arisen independently of that in the *A. varipes* complex, because these two groups also have different karyotype structures. *Aphelinus asychis* has a haploid karyotype with two metacentric chromosomes, a subtelocentric chromosome and an acrocentric chromosome, with the three smallest chromosomes being similar in size (Gokhman 2009). The same chromosome number in *A. asychis* and in the *A. varipes* complex could be an example of karyotypic orthoselection (White 1973), i.e. similar karyotypes with independent origins. However, the hypothesis of chromosomal fusion giving rise to four chromosomes in the *A. varipes* complex cannot explain the significant and substantially smaller genome sizes in this complex compared to those in the *A. mali* complex.

Chromosomes in the *A. varipes* complex differ from those in the *A. mali* complex in relative length and centromeric indices. The longest metacentric chromosome in species in the *A. varipes* complex is much longer than the other chromosomes. We suggest that this metacentric chromosome resulted from a fusion of two smaller chromosomes from an ancestral karyotype with five chromosomes. Species in the *A. varipes* complex have two smaller acrocentrics that, in turn, could originate from metacentric chromosomes of the ancestral karyotype via pericentric inversions. Moreover, the position of the centromere of the second largest chromosome underwent further changes in two sister species in the *A. varipes* complex, *A. kurdjumovi* and *A. hordei*. The centromere is significantly shifted in *A. hordei* (CI= 41 versus 46 in *A. certus*, and 47 in *A. atriplicis* and *A. varipes*), and is further moved to a terminal position in *A. kurdjumovi* (CI= 0). We propose that consecutive pericentric inversions in *A. hordei* and *A. kurdjumovi* would be the most parsimonious explanation. These chromosomal rearrangements in the *A. varipes* complex are an example of a general trend in karyotype evolution in parasitic Hymenoptera, namely, karyotypic dissymmetrization, which involves an increase in size differentiation between chromosomes and an increase in the proportion of acrocentric chromosomes (Gokhman 2009).

A recent review of the distribution of rDNA sites on chromosomes of parasitic Hymenoptera showed that the number of these sites correlates with chromosome number (Gokhman et al. 2014). We found that this is also true for at least one *Aphelinus* species: haploid males of *A. coreae*, with their low number of chromosomes ($n = 5$), had a single

rDNA site, while diploid females of *A. coreae* had two rDNA sites. Fluorescence *in situ* hybridization is especially useful for studying karyotypes with morphologically similar chromosomes that are difficult to recognize with conventional staining, like the chromosomes of *A. coreae* and other species in the *A. mali* and *A. daucicola* complexes.

Total chromosomal length was correlated with genome size in *Aphelinus*, but this was because of the difference in chromosome length between the *A. mali* and *A. varipes* complexes. Although a similar correlation was found for species in the family Figitidae (Gokhman et al. 2014), only large differences in chromosome length were distinguished in both cases, probably because of intraspecific variation in chromosomal condensation. Total lengths of male chromosomes exceeded those of females, although male and female genome sizes did not differ, and indeed it would be surprising if they did, given that males inherit their chromosomes from their mothers. The difference in the chromosome length between males and females may have resulted from differences in chromosomal condensation between the sexes, and this could compensate for the differences in chromatin available for transcription in the haploid and diploid genomes.

Conclusions

Differences as large as 44% were found in genome size between *Aphelinus* species, which is surprising given the similarity in their morphology and life history. Mean total chromosome length correlated with mean genome size. The differences in genome size and total chromosome length between species complexes matched the phylogenetic divergence between species complexes. Chromosomal rearrangements in the *A. varipes* complex are an example of karyotypic dissymmetrization, which involves an increase in size differentiation between chromosomes and an increase in the proportion of acrocentric chromosomes, which is a general trend in karyotype evolution in parasitic Hymenoptera.

Acknowledgements

We thank Kathryn Lanier and Joshua Rhoades, USDA-ARS, Newark, Delaware, for rearing the *Aphelinus* cultures. We thank Deni Galileo in the CTCR Core Facility, University of Delaware, for assistance with the flow cytometer analyses and Jeffrey Caplan and Jean Ross in the Bio-Imaging Center, Delaware Biotechnology Institute, for providing facilities and assistance with imaging of fluorescence *in situ* hybridizations. These core facilities are supported by the Delaware INBRE program, with a grant from the National Institute of General Medical Sciences grant (P20 GM103446) from the National Institutes of Health and the state of Delaware and by a National Science Foundation EPSCoR grant (IIA-1301765). This research was funded by the United States Department of Agriculture, Agriculture Research Service and by Award DEB 1257601 from the National Science Foundation to JBW and KRH and Award 15-04-07709 from the Russian Foundation for Basic Research to VEG. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References

- Ardila-Garcia AM, Gregory TR (2009) An exploration of genome size diversity in dragonflies and damselflies (Insecta: Odonata). *Journal of Zoology* 278: 163–173. <https://doi.org/10.1111/j.1469-7998.2009.00557.x>
- Ardila-Garcia AM, Umphrey GJ, Gregory TR (2010) An expansion of the genome size dataset for the insect order Hymenoptera, with a first test of parasitism and eusociality as possible constraints. *Insect Molecular Biology* 19: 337–346. <https://doi.org/10.1111/j.1365-2583.2010.00992.x>
- Ashmead WH (1888) Descriptions of some new North American Chalcididae. III: Euderinae (Hymenoptera: Chalcidoidea). *Canadian Entomologist* 20: 101–107. <https://doi.org/10.4039/Ent20101-6>
- Baur H, Kranz-Baltensperger Y, Cruaud A, Rasplus J-Y, Timokhov AV, Gokhman VE (2014) Morphometric analysis and taxonomic revision of *Anisopteromalus* Ruschka (Hymenoptera: Chalcidoidea: Pteromalidae) – an integrative approach. *Systematic Entomology* 39: 691–709. <https://doi.org/10.1111/syen.12081>
- Biemont C (2008) Within-species variation in genome size. *Heredity* 101: 297–298. <https://doi.org/10.1038/hdy.2008.80>
- Bosco G, Campbell P, Leiva-Neto JT, Markow TA (2007) Analysis of *Drosophila* species genome size and satellite DNA content reveals significant differences among strains as well as between species. *Genetics* 177: 1277–1290. <https://doi.org/10.1534/genetics.107.075069>
- Eggleton P, Belshaw R (1992) Insect parasitoids: an evolutionary overview. *Philosophical Transactions of the Royal Society of London Series B: Biological Sciences* 337: 1–20. <https://doi.org/10.1098/rstb.1992.0079>
- Evans GA, Schauff ME, Kokyokomi ML, Yokomi RK (1995) A new species of *Aphelinus* (Hymenoptera, Aphelinidae) that parasitizes the Spirea aphid, *Aphis spiraecola* Patch (Homoptera, Aphididae). *Proceedings of the Entomological Society of Washington* 97: 17–21. <https://doi.org/10.1201/b17450-8>
- Förster A (1841) Beiträge zur Monographie der Pteromalinen Nees 1 Heft:45. Aachen, 47 pp.
- Gadau J, Rütten K, Neusser M (2014) Parasitoid wasps (Hymenoptera). In: Sharakhov IV (Ed.) *Protocols for cytogenetic mapping of arthropod genomes*. CRC Press, Boca Raton, Florida 257–284.
- Gahan AB (1924) Some new parasitic Hymenoptera with notes on several described forms. *Proceedings of the United States National Museum* 65: 9.
- Galbraith DW, Harkins KR, Maddox JM, Ayres NM, Sharma DP, Firoozabady E (1983) Rapid flow cytometric analysis of the cell-cycle in intact plant-tissues. *Science* 220: 1049–1051. <https://doi.org/10.1126/science.220.4601.1049>
- Girault AA (1913) Australian Hymenoptera Chalcidoidea – IV. *Memoirs of the Queensland Museum* 2: 181–295.
- Godfray HCJ (1994) *Parasitoids: Behavioral and Evolutionary Ecology*. Princeton University Press, Princeton, New Jersey, 473 pp.
- Gokhman VE (2003) Karyotypes of parasitic Hymenoptera: evolution, systematic and phylogenetic implications. PhD dissertation, Moscow, Moscow State University, Russia, 338 pp.

- Gokhman VE (2009) Karyotypes of parasitic Hymenoptera. Springer, 183 pp. <https://doi.org/10.1007/978-1-4020-9807-9>
- Gokhman VE, Anokhin BA, Kuznetsova VG (2014) Distribution of 18S rDNA sites and absence of the canonical TTAGG insect telomeric repeat in parasitoid Hymenoptera. *Genetica* 142: 317–322. <https://doi.org/10.1007/s10709-014-9776-3>
- Gokhman VE, Johnston JS, Small C, Rajwani R, Hanrahan SJ, Govind S (2011) Genomic and karyotypic variation in *Drosophila* parasitoids (Hymenoptera, Cynipoidea, Figitidae). *Comparative Cytogenetics* 5: 211–221. <https://doi.org/10.3897/compcytogen.v5i3.1435>
- Gokhman VE, Kuznetsova VG (2006) Comparative insect karyology: current state and applications. *Entomological Review* 86: 352–368. <https://doi.org/10.1134/S0013873806030110>
- Gregory TR (2002) Genome size and developmental complexity. *Genetica* 115: 131–146. <https://doi.org/10.1023/A:1016032400147>
- Gregory TR (2005) Synergy between sequence and size in large-scale genomics. *Nature Reviews Genetics* 6: 699–708. <https://doi.org/10.1038/nrg1674>
- Gregory TR, Johnston JS (2008) Genome size diversity in the family Drosophilidae. *Heredity* 101: 228–238. <https://doi.org/10.1038/hdy.2008.49>
- Gregory TR, Nathwani P, Bonnett TR, Huber DPW (2013) Sizing up arthropod genomes: an evaluation of the impact of environmental variation on genome size estimates by flow cytometry and the use of qPCR as a method of estimation. *Genome* 56: 505–510. <https://doi.org/10.1139/gen-2013-0044>
- Haldeman SS (1851) *Eriophilus mali*. *Pennsylvania Farm Journal* 1: 130–131.
- Hanrahan SJ, Johnston JS (2011) New genome size estimates of 134 species of arthropods. *Chromosome Research* 19: 809–823. <https://doi.org/10.1007/s10577-011-9231-6>
- Hare EE, Johnston JS (2011) Genome size determination using flow cytometry of propidium iodide-stained nuclei. *Molecular Methods for Evolutionary Genetics* 772: 3–12. https://doi.org/10.1007/978-1-61779-228-1_1
- Hayat M (1972) The species of *Aphelinus* Dalman, 1820 [Hymenoptera: Aphelinidae] from India. *Entomophaga* 17: 49–58. <https://doi.org/10.1007/BF02371072>
- Hayat M (1994) Taxonomic studies on *Aphelinus* (Hymenoptera: Aphelinidae). VI. Records of two known and descriptions of two new species from the Oriental Region. *Entomon* 19: 35–39.
- Hayat M (1998) Aphelinidae of India (Hymenoptera: Chalcidoidea): a taxonomic revision. *Memoirs on Entomology, International* 13, 432 pp.
- Hayat M, Fatima K (1992) Taxonomic studies on *Aphelinus* (Hymenoptera: Aphelinidae). 5. Description of a new species and further records of *A. gossypii*, with a new synonymy. *Entomon* 17: 103–107.
- Heraty JM, Burks RA, Cruaud A, Gibson GAP, Liljeblad J, Munro J, Rasplus JY, Delvare G, Jansta P, Gumovsky A, Huber J, Woolley JB, Krogmann L, Heydon S, Polaszek A, Schmidt S, Darling DC, Gates MW, Mottern J, Murray E, Dal Molin A, Triapitsyn S, Baur H, Pinto JD, van Noort S, George J, Yoder M (2013) A phylogenetic analysis of the megadiverse Chalcidoidea (Hymenoptera). *Cladistics* 29: 466–542. <https://doi.org/10.1111/cla.12006>
- Heraty JM, Woolley JB, Hopper KR, Hawks DL, Kim JW, Buffington M (2007) Molecular phylogenetics and reproductive incompatibility in a complex of cryptic species of

- aphid parasitoids. *Molecular Phylogenetics and Evolution* 45: 480–493. <https://doi.org/10.1016/j.ympev.2007.06.021>
- Hopper KR, Woolley JB, Hoelmer K, Wu K, Qiao G-X, Lee S (2012) An identification key to species in the *mali* complex of *Aphelinus* (Hymenoptera, Chalcidoidea) with descriptions of three new species. *Journal of Hymenoptera Research* 26: 73–96. <https://doi.org/10.3897/jhr.26.2584>
- Howard LO (1908) Upon the *Aphis* feeding species of *Aphelinus*. *Entomological News* 19: 365–367.
- Imai HT, Taylor RW, Crosland MWJ, Crozier RH (1988) Modes of spontaneous chromosomal mutation and karyotype evolution in ants with reference to the minimum interaction hypothesis. *Japanese Journal of Genetics* 63: 159–185. <https://doi.org/10.1266/jjg.63.159>
- Jacobson AL, Johnston JS, Rotenberg D, Whitfield AE, Booth W, Vargo EL, Kennedy GG (2013) Genome size and ploidy of Thysanoptera. *Insect Molecular Biology* 22: 12–17. <https://doi.org/10.1111/j.1365-2583.2012.01165.x>
- Johnston JS, Ross LD, Beani L, Hughes DP, Kathirithamby J (2004) Tiny genomes and endoreduplication in Strepsiptera. *Insect Molecular Biology* 13: 581–585. <https://doi.org/10.1111/j.0962-1075.2004.00514.x>
- Kim JW, Heraty J (2012) A phylogenetic analysis of the genera of Aphelininae (Hymenoptera: Aphelinidae), with a generic key and descriptions of new taxa. *Systematic Entomology* 37: 497–549. <https://doi.org/10.1111/j.1365-3113.2012.00625.x>
- Kurdjumov NV (1913) Notes on European species of the genus *Aphelinus* Dalm. (Hymenoptera, Chalcidoidea), parasitic upon the plant-lice. *Russian Review of Entomology* 13: 266–270.
- Levan A, Fredga K, Sandberg AA (1964) Nomenclature for centromeric position on chromosomes. *Hereditas* 52: 201–220. <https://doi.org/10.1111/j.1601-5223.1964.tb01953.x>
- Lopes DM, de Carvalho CR, Clarindo WR, Praca MM, Tavares MG (2009) Genome size estimation of three stingless bee species (Hymenoptera, Meliponinae) by flow cytometry. *Apidologie* 40: 517–523. <https://doi.org/10.1051/apido/2009030>
- Macgregor HC, Varley JM (1988) Working with animal chromosomes. John Wiley & Sons, Chichester, 290 pp.
- Maddison WP, Maddison DR (2016) Mesquite: a modular system for evolutionary analysis. Version 3.11 <http://mesquiteproject.org>.
- Matsumoto K, Yamamoto DS, Sumitani M, Lee JM, Hatakeyama M, Oishi K (2002) Detection of a single copy gene on a mitotic metaphase chromosome by fluorescence in situ hybridization (FISH) in the sawfly, *Athalia rosae* (Hymenoptera). *Archives of Insect Biochemistry and Physiology* 49: 34–40. <https://doi.org/10.1002/arch.10005>
- Nikol'skaya MN, Yasnosh VA (1966) Aphelinidae of the European Part of the USSR and Caucasus (Chalcidoidea: Aphelinidae). Nauka, Leningrad, 295 pp.
- Pan MX (1992) A new species of *Aphelinus* Dalman from Zhejiang province, China (Hymenoptera: Aphelinidae). *Acta Zootaxonomica Sinica* 17: 75–77 [In Chinese].
- Pinkel D, Straume T, Gray JW (1986) Cytogenetic analysis using quantitative, high-sensitivity, fluorescence hybridization. *Proceedings of the National Academy of Sciences of the United States of America* 83: 2934–2938. <https://doi.org/10.1073/pnas.83.9.2934>

- Prinsloo GL, Naser OC (1994) The southern African species of *Aphelinus* Dalman (Hymenoptera: Aphelinidae), parasitoids of aphids (Homoptera: Aphidoidea). *Journal of African Zoology* 108: 143–162.
- R_Core_Team (2014) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <http://www.R-project.org/>
- Rice A, Glick L, Abadi S, Einhorn M, Kopelman NM, Salman-Minkov A, Mayzel J, Chay O, Mayrose I (2014) The Chromosome Counts Database (CCDB) – a community resource of plant chromosome numbers. *New Phytologist* <http://ccdb.tau.ac.il/> [accessed 24 January 2015]
- Ross L, Blackmon H, Lorite P, Gokhman VE, Hardy NB (2015) Recombination, chromosome number and eusociality in the Hymenoptera. *Journal of Evolutionary Biology* 28: 105–116. <https://doi.org/10.1111/jeb.12543>
- Rössler Y, DeBach P (1973) Genetic variability in a thelytokous form of *Aphytis mytilaspidis* (LeBaron) (Hymenoptera: Aphelinidae). *Hilgardia* 42: 149–175. <https://doi.org/10.3733/hilg.v42n05p149>
- Sharakhova MV, George P, Timoshevskiy V, Sharma A, Peery A, Sharakhov IV (2014) Mosquitoes (Diptera). In: Sharakhov IV (Ed.) *Protocols for cytogenetic mapping of arthropod genomes*. CRC Press, Boca Raton, Florida, 93–170. <https://doi.org/10.1201/b17450-5>
- Timberlake PH (1924) Descriptions of new Chalcid-flies from Hawaii and Mexico (Hymenoptera). *Proceedings of the Hawaiian Entomological Society* 5: 395–417.
- Vergilino R, Dionne K, Nozais C, Dufresne F, Belzile C (2012) Genome size differences in *Hyalella* cryptic species. *Genome* 55: 134–139. <https://doi.org/10.1139/g11-085>
- Viggiani G (1967) *Ricerca sugli Hymenoptera Chalcidoidea*. XV. Osservazioni carilogiche preliminari sull'*Aphelinus mali* (Hald.). *Bolletino del Laboratorio di Entomologia Agraria "Filippo Silvestri"* 25: 326–330.
- White MJD (1973) *Animal Cytology and Evolution*. Cambridge University Press, Cambridge, 961 pp.
- Yasnosh VA (1963) New species of the genus *Aphelinus* Dalm. (Hymenoptera, Chalcidoidea) in the fauna of the USSR. *Entomological Review* 42: 178–189.
- Zehavi A, Rosen D (1988) A new species of *Aphelinus* (Hymenoptera: Aphelinidae) from Israel, with notes on the mali group. *Israel Journal of Entomology* 22: 101–108.